DEPLETION OF Mg²⁺ AND PERMEABILITY INCREASE OF THE MITOCHONDRIAL INNER MEMBRANE BY PRIMYCIN

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Recently we have shown¹⁾ that primycin below 2~3 nmoles/mg protein concentration ("low dose") renders their inner membrane permeable to K⁺, Na⁺ and Tris⁺ but not to Tea⁺. This effect was seen only under energized conditions. It has been concluded that this effect of the antibiotic is due to its ionophore-like action. This conclusion has been supported by the experiments of BLASKÓ et al.2) made on erythrocytes and artificial lipid membranes. On the other hand, we have also reported¹⁾ that primycin at a concentration higher than 2~3 nmoles/mg protein ("high dose") can also interact with nonrespiring (i.e. deenergized) mitochondria increasing their inner membrane permeability to protons and chloride, too. Since the ATPase activity induced by a "high dose" of primycin was significantly higher in the presence of added Mg²⁺ than in its absence, the depletion of Mg²⁺ from the mitochondria by the antibiotic has been proposed to be responsible for the non-selective permeability increase. In this note we present direct evidence in favour of this proposal.

As it can be seen in Fig. 1 primycin in a "high dose" (6.6 nmoles/mg protein) induces a rapid and in the presence of EDTA practically complete —depletion of Mg^{2+} from mitochondria, while adding it in "low dose" (1 nmole/mg protein) the Mg^{2+} loss is significantly less and slower.

Our previous proposal that primycin in a "high dose" depletes Mg^{2+} from mitochondria is now experimentally verified. Thus the non-

Fig. 1. Mg²⁺ and Ca²⁺ efflux from mitochondria induced by primycin.

The media contained 140 mm sucrose, 120 mm mannitol, 2 mM HEPES, 2.5 mM succinate and 5 µM rotenone. The pH of reagents and the media was adjusted to 7.4 with KOH. Further additions were: •—• none, 0—0 0.5 mм EDTA, —— 1 nmole of primycin/mg protein alone,
plus EDTA, A-A 6.6 nmoles of primycin/mg protein alone, $\triangle - \triangle$ plus EDTA. The reaction was started at 25°C by addition of mitochondria prepared by the method of JOHNSON and LARDY¹⁰⁾, and was stopped by cooling the samples to 0°C and by centrifugation (2 min, $16,000 \times g$). The amount of bivalent cations was measured by atomic absorption spectrometry following the method of REED and LARDY¹¹⁾. The mitochondrial protein content of all tubes was 5.4 mg determined according to SCHACTERLE and POLLACK¹²⁾.



selective permeability changes can be well explained by the depletion of Mg^{2+} from mitochondria as suggested by others^{3~7}.

To see whether primycin in a "high dose" also induces a Mg^{2+} influx, swelling experiments were made in isoosmotic $Mg(NO_3)_2$. Figs. 2 a, b, c show that in the presence of "low dose" of the antibiotic neither energized nor deenergized mitochondria swell in magnesium nitrate, though they do swell in isoosmotic KNO_3 . On the contrary, primycin in a "high dose" induces a high amplitude swelling of non-respiring mitochondria in magnesium nitrate (Fig. 2a). Thus it can be concluded that primycin only in a "high dose" renders the membrane permeable to Mg^{2+} .

The question arises whether primycin in "high dose" acts as a bivalent cationophore.

As it is shown in Fig. 1 primycin in "low" and "high dose" equally induces a Ca^{2+} efflux from mitochondria. The rate of this efflux is increased in the presence of EDTA presumably preventing the mitochondrial reuptake of Ca^{2+} .

Non-respiring mitochondria swell in isoos-

Abbreviations: Tris, Tris (hydroxymethyl) aminomethane; Tea, triethanolamine; EDTA, ethylenediaminetetraacetic acid; HEPES, N-2-hydroxyethylpiperazine-N'-2-ethanesulphonic acid; CCCP, carbonyl cyanide *m*-chlorophenylhydrazone.

Fig. 2. Osmotic swelling of mitochondria induced by primycin in isoosmotic solutions.

The swelling was followed monitoring the changes of optical density at 610 nm. The medium contained 100 mM Mg(NO₃)₂ in a, b; 140 mM KNO₃ in c and 100 mM Ca(CH₃COO)₂ in d, e, f. 5 mM glutamate and 1.7 mM malate were also present in a, b and c. The pH was adjusted to 7.4 with 5 mM Tea-NO₃ in a, b, c and with acetic acid in d, e and f. The amount of mitochondrial protein was 3.9 mg in each experiment. Other additions at the arrows were: rotenone (3 μ M Rot) in d, e, f; CCCP (3 μ M); ruthenium red (RR 6 nmoles) and primycin. The numbers by the curves of a, b, c represent the concentration of antibiotic expressed in nmoles/mg protein. The concentration of primycin in d, e and f was 6.6 nmoles/mg protein.



motic calcium acetate in the presence of protonophore, and this swelling can be prevented by ruthenium red which inhibits the natural Catranslocator of the inner membrane (Figs. 2d and e). The synthetic Ca-ionophore, as found by CARONI *et al.*⁹, induces a swelling of mitochondria also in the presence of ruthenium red, by facilitating the Ca²⁺ influx through the inner membrane. On the contrary, primycin does not induce a swelling of ruthenium red treated mitochondria indicating that it does not facilitate the transport of Ca²⁺ (Figs. 2e and f).

These experimental findings make very unlikely that primycin in "high dose" acts as a bivalent cationophore. Thus, the mechanism by which the antibiotic increases the Mg^{2+} permeability of the inner membrane of mitochondria requires further experiments. In this respect it should be mentioned, that oleficin, an antibiotic of quite different structure, increases the Mg^{2+} permeability of the mitochondrial inner membrane in a similar manner⁹⁾.

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